# ORIGINAL ARTICLE



# Molecular phylogeny of the endemic fern genera Cyrtomidictyum and Cyrtogonellum (Dryopteridaceae) from East Asia

Hong-Mei Liu · Xian-Chun Zhang · Wei Wang · Hui Zeng

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**Abstract** Cyrtomidictyum Ching and Cyrtogonellum Ching are two eastern Asian endemic genera whose taxonomic affinities and phylogenetic relationships have long been controversial. The main uncertainty surrounds the separation of the two genera from the species-rich genus Polystichum. Here we present a phylogenetic study focusing on the phylogenetic relationships of these polystichoid ferns. We reconstructed the relationships based on DNA sequence variation in four chloroplast genome regions, rbcL, atpB, and the intergenic spacers (IGS) rps4-trnS and trnL-trnF. Maximum likelihood and Bayesian inference analyses confirm earlier results that were based on less comprehensive taxon sampling and either only a single gene (rbcL) or two IGS (rps4-trnS and trnL-trnF). Cyrtomidictyum is the sister of the clade of polystichoid ferns that includes Cyrtogonellum, Cyrtomium subser. Balansana and three sections of Polystichum. Cyrtogonellum groups with several species of Polystichum, and constitutes the sister taxon to Polystichum sect. Sphaenopolystichum. We support the recognition of Cyrtomidictyum as circumscribed initially, rather than expansion of the genus to include either several Polystichum species or Cyrtogonellum, some Polystichum and Cyrtomium species. The monophyly of Cyrtomidictyum is supported by morphological characters such as once-pinnate leaves, free venation, prolongated leaf apices, and exindusiate sori. Two synapomorphic indels in the chloroplast genome, one 15-bp deletion in *rps4-trnS*, and one 3-bp insertion in *trnL-trnF* further differentiate *Cyrtomidictyum* from other polystichoid ferns. The close affinity of *Cyrtogonellum* to section *Sphaenopolystichum* of *Polystichum* s.s. is highly supported by molecular data. However, no shared morphological characters or molecular indels have been detected, although the distinctness of *Cyrtogonellum* is shown by a 13-bp insertion in the *rps4-trnS* alignment.

**Keywords** East Asia · Molecular phylogeny · atpB · rbcL · rps4-trnS · trnL-trnF · IGS

# Introduction

Polystichum Roth is one of the ten most species-rich genera of ferns and has its main centre of diversity in southwest China and adjacent regions (Kung et al. 2001). Several small genera with restricted distribution ranges—Acropelta Nakai, Cyrtomium C. Presl, Cyrtomidictyum Ching, Cyrtogonellum Ching, Papuapteris C. Chr., Phanerophlebia C. Presl, Plecosorus Fée, Ptilopteris Hance, and Sorolepidium Christ emend. Ching—have been suggested to be included within Polystichum (Kramer et al. 1990; Tryon and Tryon 1982). Recent molecular evidence found the broadly defined Polystichum to be non-monophyletic (Li et al. 2008; Little and Barrington 2003; Lu et al. 2007). Among several putative segregates sampled, Cyrtomium, Phanerophlebia and Cyrtomidictyum were recognized as separate genera within Polystichum s.l., and a narrower, monophyletic Polystichum was accepted (Li et al. 2008; Little and Barrington 2003; Lu et al. 2007). Chloroplast DNA-based phylogenetic analyses led to the additional inclusion in

H.-M. Liu · H. Zeng (⊠)

The Key Laboratory for Environmental and Urban Sciences, Shenzhen Graduate School, Peking University, Shenzhen 518055, People's Republic of China e-mail: zengh@szpku.edu.cn

X.-C. Zhang · W. Wang State Key Laboratory of Systematic and Evolutionary Botany, Institute of Botany, Chinese Academy of Sciences, Beijing 100093, People's Republic of China

*Polystichum* s.s. of *Plecosorus* (Little and Barrington 2003) and *Sorolepidium* (Liu et al. 2007a; Lu et al. 2007).

Cyrtogonellum is an endemic genus of polystichoid ferns including eight described species mainly distributed in limestone areas of Guizhou and adjacent regions, with two species extending to northern Vietnam (Wu et al. 2005; Xie 1990; Fig. 1). Ching (1938) proposed Cyrtogonellum as morphologically intermediate between Cyrtomium and Polystichum, but considered the genus as more closely related to Phanerophlebia on account of similar growth habit, venation and leaf texture. More recently, Cyrtogonellum was usually regarded as more closely related to Polystichum (Ching 1978; Tryon and Tryon 1982; Xie 1990). Several non-Chinese authors considered the morphological evidence as insufficient support for Cyrtogonellum as a distinct genus and treated it as a synonym of Phanerophlebia (Copeland 1947), Cyrtomium (Tryon and Lugardon 1991) or *Polystichum* (Kramer et al. 1990).

Two or three *Cyrtogonellum* species were included in recent phylogenetic analyses of polystichoid ferns, within which a monophyletic clade ("BCPC" of Lu et al. 2007; "CCCP" of Li et al. 2008) comprising *Cyrtomidictyum*, *Cyrtogonellum*, *Cyrtomium* subser. *Balansana* and three sections of *Polystichum* was recognized. However, the exact relationships among these taxa were not resolved. *Cyrtogonellum* grouped with several species assigned to *Polystichum* in phylogenetic analysis of the family Dryopteridaceae based on the chloroplast genes *rbc*L and *atp*B (Liu et al. 2007b). However, the two included species of *Cyrtogonellum* did not group together but appeared on divergent branches, although this part of the topology was poorly supported (less than 50% BS).

Cyrtomidictyum is another small polystichoid genus; it comprises four species (Ching 1940, 1957). The genus is mainly distributed in eastern and southeastern China, but

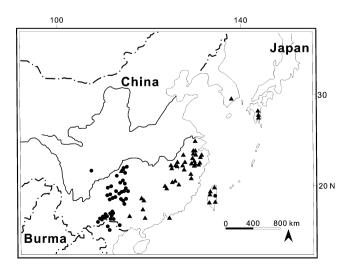


Fig. 1 Geographical distribution of *Cyrtomidictyum* (triangles) and *Cyrtogonellum* (circles)



the range of the type species, C. lepidocaulon, also extends to Japan and South Korea (Fig. 1). Cyrtomidictyum was accepted as a distinct genus in some recent classifications (Ching 1978; Pichi Sermolli 1977; Tryon and Tryon 1982) but not by Kramer et al. (1990) who included it in a more broadly defined Polystichum. In a recent worldwide phylogeny of *Polystichum*, Little and Barrington (2003) considered Cyrtomidictyum as remotely related to Polystichum s.s. and suggested its recognition as a distinct genus. These authors also proposed the expansion of Cyrtomidictyum to include two Polystichum species, P. tripteron and P. deltodon. Subsequent phylogenetic study of Chinese Polystichum confirmed the separation of Cyrtomidictyum from Polystichum s.s., suggesting expansion of the genus Cyrtomidictyum to include Cyrtogonellum, some species of Cyrtomium and Polystichum, namely the CCPC clade (Li et al. 2008). Recently, using rbcL, Lu et al. (2007) considered the BCPC clade as a distinct genus comprising Cyrtomidictyum, Cyrtogonellum (three representatives), Cyrtomium subser. Balansana, and three sections of Polystichum (sect. Haplopolystichum, sect. Sphaenopolystichum, sect. Crucifilix). Although no final taxonomic treatment was provided, it was proposed to use the generic name of Cyrtogonellum, which had been published two years prior to Cyrtomidictyum. However, only one of the four species of Cyrtomidictyum was sampled and the respective studies used only rbcL (Little and Barrington 2003; Lu et al. 2007) or the rps4-trnS and trnL-trnF spacers (Li et al. 2008). In summary, the exact generic circumscriptions and phylogenetic relationships among Cyrtomidictyum, Cyrtogonellum, Cyrtomium and Polystichum were not fully understood or at least required confirmation using additional taxon sampling.

In the present study, we generated sequence data from four chloroplast genes, the two coding regions rbcL and atpB as well as the trnL-trnF and rps4-trnS IGS regions, for the first time including rps4 and rps4-trnS IGS for all species of Cyrtomidictyum. In addition, we combined sequences for Cyrtogonellum from our previously published dataset (Liu et al. 2007a, b) and other published studies (Li et al. 2008; Lu et al. 2007). The comprehensive taxon sampling allows us to address the main objective of this study, i.e. generic circumscriptions and phylogenetic relationships of these two small genera, by adding new evidence based on exhaustive taxon sampling and multiple gene analyses.

# Material and methods

Taxon sampling and sources of sequences

A total of 53 polystichoid samples was included in the study group. The ingroup comprised four *Cyrtomidictyum* 

species and five species of *Cyrtogonellum*, plus representatives of two subgroups of *Cyrtomium* as recognized by Lu et al. (2005) and 34 species of *Polystichum* representing all sections of Kung et al. (2001).

We obtained nine new sequences for *Cyrtomidictyum* species: *C. conjunctum*: *atp*B (EF540725), *rps*4-*trn*S (EF540711); *C. basipinnatum*: *rbc*L (EF463122), *atp*B (EF450478), *rps*4-*trn*S (EF540712); *C. faberi*: *rbc*L (EF463124), *atp*B (EF450479), *rps*4-*trn*S (EF540710), *trn*L-*trn*F (EF540697). The corresponding voucher specimens have been deposited in the Herbarium of the Institute of Botany, Chinese Academy of Sciences, Beijing (PE). Taxon information and related references are listed in Table 1.

RbcL sequences were available for all included taxa, but sequences of the rps4-trnS and trnL-trnF IGS were missing for some species. Sequences of trnL-trnF were missing for Cyrtomidictyum basipinnatum and C. conjunctum, Cyrtogonellum xichouensis and Polystichum tonkinense, of rps4trnS for Cyrtogonellum caducum, C. fraxinellum, C. inaequalis and Cyrtomium fortunei. A combined data set was collected including 18 taxa, 14 polystichoid ferns and four outgroup taxa. For the latter species all four chloroplast genome sequences were available, except that atpB sequences were lacking for three species. All missing sequences were treated as 'missing data' in the phylogenetic analyses. No sequence data exist for rps4-trnS and trnL-trnF in Phanerophlebia and Polystichopsis. Therefore, four species of Arachniodes and Dryopteris were selected as outgroup taxa. The different species of Arachniodes were lumped into a single outgroup taxon, with the rbcL, atpB and rps4-trnS data deriving from A. rhomboidea, trnL-trnF from A. aristata.

# DNA extraction, PCR amplification and sequencing

Silica gel-dried leaf material was newly obtained for Cyrtomidictyum faberi, C. conjunctum and C. basipinnatum, and for several species of Polystichum. Total genomic DNA was extracted using the 2×CTAB method adapted from Doyle and Doyle (1987). The polymerase chain reaction (PCR) was used to amplify four chloroplast genes or spacer regions. Amplification and sequencing of the rbcL gene was carried out using the forward primer 1F (Chen et al. 1998) and the reverse primer 1351R (Gastony and Rollo 1995), with the additional internal sequencing primer 440F (Gastony and Rollo 1995). The atpB gene was amplified and sequenced using forward primer F and reverse primer R as described by Liu et al. (2007b). Amplification and sequencing of the rps4-trnS region was accomplished with forward primer F and reverse primer R (Thomson et al. 2005), for trnL-trnF with forward primer C and reverse primer F (Taberlet et al. 1991). The resulting PCR products were purified using a GFX<sup>TM</sup> PCR DNA and Gel Band Purification Kit according to the manufacturer's instructions, and then directly sequenced. Sequencing reactions were conducted using the DYEnamic<sup>TM</sup> ET Dye Terminator Cycle Sequencing Kit. Sequences were analyzed using MegaBACE<sup>TM</sup>1000 DNA Analysis Systems (Amersham Biosciences), following the manufacturer's protocol.

Sequence alignment, indel coding, and phylogenetic analyses

Alignments of all sequences were generated using CLUS-TAL X (Thompson et al. 1997) and subsequently edited manually in BioEdit (Hall 1999) for the IGS regions of rps4-trnS and trnL-trnF. The rbcL and atpB sequence alignments did not include insertions or deletions. Indels were required to be introduced into the alignment of the rps4-trnS and trnL-trnF spacer regions. Indels shared by two or more taxa were treated as potentially phylogenetically informative characters. Ambiguously aligned regions were excluded from all subsequent analyses. Phylogenetic analyses were conducted using both Maximum Likelihood (ML) and Bayesian Inference of phylogeny (BI). Maximum Likelihood analyses were conducted using PHYML version 2.4.3 (Guindon and Gascuel 2003) under the optimal model of sequence evolution. For each data set, an Akaike-Information-Criterion as implemented in Modeltest 3.07 (Posada and Crandall 1998) was used to select the model of nucleotide substitution, including gamma shape and proportion of invariant sites. Maximum Likelihood bootstrap support (BS) values were estimated from 1,000 replicates in PHYML. Bayesian Inference was performed with MrBayes version 3.0b4 (Ronquist and Huelsenbeck 2003). We used uniform prior probabilities and a random starting tree. Models and corresponding parameters were selected using the Akaike-Information-Criterion as implemented in Modeltest (Posada and Crandall 1998). Markov chains were run for 1,000,000 generations and sampled every 100 generations. We ran four Markov Chain Monte Carlo chains, sampling every 100 generations for 1,000,000 generations, starting with a random tree. Consensus trees of the halfcompat and allcompat types were created from the saved trees. Stationarity was reached at approximately generation 5,000; thus, the first 50 trees were the 'burnin' of the chain, and phylogenetic inferences are based on those trees sampled after generation 5,000. Excluding the burn-in, all trees sampled from the four analyses were combined to produce a consensus tree, which formed the basis for calculation of Bayesian posterior probabilities (PP) for the respective clades.

Various data matrices were used in the phylogenetic analyses: 1) *rbc*L; 2) *atp*B; 3) *rps4-trn*S; 4) *trnL-trn*F; 5)



Iable 1         Genbank accession numbers for sequences used in	s accession numb	ers ror sequence		corresponding	voucner specimens	(provided only for	this study, and corresponding voucner specimens (provided only for sequences not used in previous publications)	previous publication	ns)
Species	Voucher	rbcL	Reference	atpB	Reference	rps4-trnS	Reference	trnL-trnF	Reference
Ingroup									
C. caducum	H. M. Liu	EF463120*		EF450476*		Missing		AY736350	Lu et al. (2005)
C. falcilobum	GX273	DQ054514	Li and Lu	missing		DQ202440	Li	DQ202409	Li (in GenBank)
C. fraxinellum	X. C. Zhang	EF463121*	(2006)	EF450477*		missing	(in GenBank)	AY736349	Lu et al. (2005)
C. inaequalis	2596	AY694812	Lu et al.	missing		missing		AY736351	Lu et al. (2005)
C. xichouense		DQ054515	(2005) Li and Lu	missing		DQ202441	Li (in GenRant)	missing	
Cyrtomidictyum			(0007)				(III OGIIBAIIK)		
C. basipinnatum	C. H. Li	EF463122*		EF450478*		EF540712*		missing	
C. conjunctum	X. C. Zhang	EF463123*		EF540725*		EF540711*		missing	
C. faberi	H. Zhang	EF463124*		EF450479*		EF540710*		EF540697*	
C. lepidocaulon	BY03	DQ508767	Lu et al. (2007)	EF450480	Liu et al. (2007b)	DQ151855	Li (in GenBank)	DQ514489	Lu et al. (2005)
Cyrtomium									
C. aequibasis		AY694809	Lu et al. (2005)	ı		DQ202442	Li (in GanBank)	AY736346	Lu et al. (2005)
C. balansae		AY694799	Lu et al. (2005)	missing		DQ202443	(III Gelibalik) Li	AY736335	Lu et al. (2005)
C. caryotideum		AF537225	Little and	ı		DQ202444	(in GenBank) Li	AY736347	Lu et al. (2005)
C. devexiscapulae		AY694798	Barrington (2003) Lu et al. (2005)	I		DQ202445	(m GenBank) Li	AY736334	Lu et al. (2005)
C. fortunei		EF39423	Liu et al. (2007a)	EF450471	Liu et al.	missing	(in Genbank)	AY736348	Lu et al. (2005)
C. hookerianum		AY694801	Lu et al. (2005)	EF540726	(2007b) Liu et al.	DQ202446	Li	AY736337	Lu et al. (2005)
C. uniseriale		AY 694794	Lu et al. (2005)	I	(9/007)	I	(III Genbank)	I	
C. urophyllum		EF394240	Liu et al. (2007a)	ı		ı		ı	
Phanerophlebia		AF537233	Little and	I		I		ı	
umbonata Polystichonsis		AF537234	Barrington (2003)	ı		ı		I	
chaerophylloides Polystichum			Barrington (2003)	ı		1		ı	
P. acutidens		EF394244	Liu et al. (2007a)	EF450459	Liu et al.	DQ202451	Li (in GenRank)	DQ202419	Li (in GenBank)
P. alfarii		AF537236	Little and Barrington (2003)	ı	(01007)	ı	(III OVIIDAIIN)	ı	
P. ammifolium		AF537237	(2003) Little and Barrington (2003).	I		I		ı	



4	P. anomalum		EF394248	Liu et al. (2007a)	ı		1		ı	
7	P. atkinsonii		EF394250	Liu et al. (2007a)	ı		ı		I	
7	P. attenuatum		AY545503	Li et al. (2004)	1		1		I	
7	P. brachypterum		AY545501	Li et al. (2004)	ı		DQ202452	Li (in GenBank)	DQ202420	Li (in GenBank)
7	P. capillies		EF394249	Liu et al. (2007a)	ı		1		ı	
7	P. christii		AY545486	Li et al. (2004)	missing		DQ151862	Li (in GenBank)	DQ150399	Li (in GenBank)
,	P. craspedosorum	X.C. Zhang	AF537238	Little and	EF463139*		DQ202454	Li (in GenBank)	DQ202422	Li (in GenBank)
,	P. crinigerum	3389	AY694813	Barrington (2003) Lu et al. (2005)	I		DQ202455	Li (in GenBank)	DQ202423	Li (in GenBank)
7	P. deltodon		AF537239	Little and	missing		DQ202456	Li (in GenBank)	DQ202424	Li (in GenBank)
.7	P. duthiei		EF394252	Barrington (2003) Liu et al. (2007a)	I		ı		ı	
1	P. ekmanii		AF537242	Little and	ı		I		I	
	P. grandifrons		AY545484	Barrington (2003) Li et al. (2004)	I		DQ202459	Li (in GenBank)	DQ202427	Li (in GenBank)
7	P. hancockii		EF394243	Liu et al. (2007a)	I		ı		I	
7	P. lachenense		EF394251	Liu et al. 2007a, b	I		I		I	
•	P. lehmannii		AF537245	Little and Barrington	I		ı		ı	
7	P. lentum		AF537246	(2003) Little and Barrington	ı		ı		ı	
7	P. lonchitis		AF537247	(2003) Little and Barrington	I		I		I	
7	P. luctuosum		EF394245	(2003) Liu et al. (2007a)	EF45046	Liu et al.	DQ151881	Li (in GenBank)	DQ150419	Li (in GenBank)
7	P. makinoi		EF394247	Liu et al. (2007a)	EF450467	(2007b) Liu et al.	DQ202462	Li (in GenBank)	DQ202431	Li (in GenBank)
	P. neolobatum		EF394246	Liu et al. (2007a)	EF450468	(2007b) Liu et al. (2007b)	DQ202463	Li (in GenBank)	DQ202432	Li (in GenBank)
7	P. nepalense		AY545499	Li et al. (2004)			DQ202465	Li (in GenBank)	AY534748	Li (in GenBank)
-	P. omeiense		AY545497	Li et al. (2004)	EF450469	Liu et al. (2007b)	DQ202466	Li (in GenBank)	DQ202434	Li (in GenBank)
	P. speciosissimum		AF537255	Little and Barrington (2003)	I		ı			
	P. stenophyllum		AF537256	Little and Barrington (2003)	DQ202471	Li (in GenBank)	ı		DQ202439	Li (in GenBank)
	P. subacutidens		AY545488	Li et al. (2004)	DQ151880	Li (in GenBank)	ı		DQ514518	Lu and Li (in GenBank)
	P. tonkinense		EF540724	Liu et al. (2007b)	EF540728	Liu et al. (2007b)	EF540714*		missing	
7	P. tripteron		U30832	Hasebe et al. (1995)	I		I		ı	
-	P. vestitum		AF208395	Perrie (in GenBank)	ı		I		ı	
-	P. xiphophyllum		DQ508788	Lu et al. (2007)	ı		DQ151882	Li (in GenBank)	DQ150420	Li (in GenBank)
,	P. yuanum		AY545487	Li et al. (2004)	ı		ı		ı	
, -	P. yunnanense		AY545504	Li et al. (2004)	ı		ı		ı	



Table 1 (continued)									
Species	Voucher	rbcL	Reference	atpB	Reference	rps4-trnS	Reference	trnL-trnF	Reference
Arachniodes									
A. caudata		EF463110	Liu et al. (2007b)	EF450482	Liu et al.	DQ191889	Li (in GenBank)	AY736356	Lu et al. (2005))
A. rhomboidea		EF463115	Liu et al. (2007b)	EF450487	(2007b) Liu et al. (2007b)	(A. giobisora) DQ191890	Li and Lu (2006)	(A. tonkmensts) AY268782 (A. aristata)	Geiger and Ranker
Dryopteris									
D. bissetiana		AY268862	Geiger and Ranker (2005)	EF450490	Liu et al. (2007b)	DQ191829	Li and Lu (2006)	AY268796	Geiger and Ranker (2005)
D. cycadina		EF463127	Liu et al. (2007b)	EF450492	Liu et al. (2007b)	DQ191835	Li and Lu (2006)	AY278400	Geiger and Ranker (in GenBank)

Newly generated sequences marked by asterisks (\*); for previously published sequences, references are given A few congeneric species were combined to composite taxa in the combined data matrix

rbcL+rps4-trnS+trnL-trnF; 6) rbcL+atpB+rps4-trnS+trnL-trnF. When separate analyses of each gene/spacer region did not result in topological conflicts among major clades supported by BS values greater than 70%, the partitions were considered as congruent and were combined in subsequent analyses.

#### Results

Analyses of the rbcL gene data

The data set included 53 taxa and 1204 characters (Fig. 2). The ML analysis of the rbcL data resulted in a topology (not shown) congruent with that from the BI analysis. With Phanerophlebia and Polystichopsis as outgroup, all remaining taxa of polystichoid ferns were resolved in two main clades: clade I comprising Polystichum s.s. and Cyrtomium s.s.; clade II including Cyrtomidictyum, Cyrtogonellum, Cyrtomium subser. Balansana and members of three sections of Polystichum. Cyrtomidictyum was resolved as a monophyletic genus with strong support (BS 100%; PP= 1.00), and as sister to the remaining members of the strongly supported clade II. Cyrtogonellum was resolved as sister group to Polystichum sect. Sphaenopolystichum. This relationship was not supported in the ML analysis (BS<50%), while the BI analysis resulted in strong support (PP=1.00).

# Analyses of the three-gene data

The three-gene data matrix included 39 accessions for rbcL, rps4-trnS and trnL-trnF, and contained 2,056 nucleotide sites. Separate analyses of the three cpDNA markers did not reveal topological conflicts among the major clades (Figs. 3, 4); thus, the partitions were considered as congruent and were combined in subsequent analyses. Analysis of the three-gene data set using Maximum Likelihood resulted in a tree topology (not shown) nearly identical to the Bayesian tree. Inferences from the three-gene combined data set supported the polystichoid fems as a monophyletic lineage (BS 100%; PP=1.00; Figs. 3, 4) which is further supported by the shared presence of indels "a", "b", "c" and "h" in the rps4-trnS spacer region (Fig. 4; Table 2). Polystichum s.l. and Cyrtomium s.l. were not monophyletic, as respective members were found in different clades, whereas monophyly of Polystichum s.s. and Cyrtomium s.s. was clearly evident and strongly supported in all analyses (Figs. 3, 4), as well as from the shared presence of insertions "o" and "n", respectively (Fig. 4; Table 2). Using Bayesian inference, Cyrtomidictyum was monophyletic and resolved as sister to the clade comprising Cyrtogonellum, Cyrtomium subser. Balansana, and sect. Haplopolystichum and sect. Sphaenopolystichum of

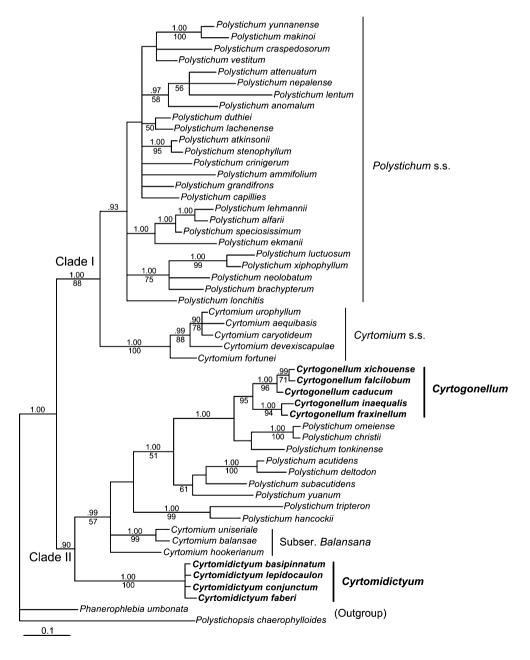


Polystichum. The distinction of Cyrtomidictyum was supported by insertion "m" in trnL-trnF and deletion "i" close to the trnS region, with the exception of C. lepidocaulon (Fig. 4; Table 2). Cyrtogonellum was supported as a monophyletic group nested in the genus Polystichum based on the combined three-gene data set. Its distinctness was supported by the unique insertion "d" in rps4-trnS (Fig. 4; Table 2). Close relationships among Cyrtogonellum, Polystichum sect. Sphaenopolystichum and sect. Haplopolystichum, Cyrtomium subser. Balansana, and Cyrtomidictyum (namely members of clade II in the rbcL tree) are further supported by the synapomorphic insertion "g" near the trnS region.

Analyses of the four-gene data

The data set consisting of four different regions in general gave greater support than analyses of both the single-gene and the 3-gene data sets (Fig. 5). *Cyrtogonellum* was supported as a monophyletic lineage and was nested in *Polystichum* s.s. with high support values (BS 100%; PP=1.00). The sister-taxon relationship of *Cyrtogonellum* to *Polystichum* sect. *Sphaenopolystichum* was resolved and strongly supported, with *Cyrtomium* subser. *Balansana* and *Polystichum* sect. *Haplopolystichum* as successive sisters to this *Cyrtogonellum-Sphaenopolystichum* lineage.

Fig. 2 50% majority rule consensus tree from four Bayesian analyses of 1,000,000 generations each of the *rbc*L data set, excluding burn-in trees. Posterior probability support values≥0.90 from Bayesian analyses shown above branches, bootstrap support values>50% from Maximum Likelihood analyses below branches





Cyrtomidictyum (C. lepidocaulon) was sister to all abovementioned groups (Fig. 5).

Polystichum craspedosorum was found in conflicting positions within the polystichoid lineage when different data sets were analysed. In the *rbcL* analysis *P. craspedosorum* grouped in *Polystichum* s.s., although support for this was not high (BS 50%; PP=0.84). In both the *rps4-trn*S and *trnL-trn*F data sets, *P. craspedosorum* was closely related to the *Cyrtomidictyum-Cyrtogonellum* clade, with high support values (Fig. 3). *Atp*B data gave only low support values for this relationship (BS<50%; PP<0.5). The shared five base-pair insertion (TCTCT) in the *rps4-trn*S region supported the close affinity of *P. craspedosorum* to the *Cyrtomidictyum-Cyrtogonellum* clade.

#### Discussion

The phylogenetic analyses of multiple chloroplast genes of polystichoid ferns confirmed previously published results and indicated that *Polystichum* and *Cyrtomium* as tradi-

tionally recognized are not monophyletic (Li et al. 2008; Little and Barrington 2003; Liu et al. 2007b; Lu et al. 2007). The monophyly of *Polystichum* s.s. was supported by insertions "o" in the *trnL-trnF* and "f" in the *rps4-trnS* region; the distinction of *Cyrtomium* s.s. was supported by four indels, three ("e", "j", "k") in the *rps4-trnS*, and one ("n") in the *trnL-trnF* region.

Generic circumscription and systematic position for *Cyrtomidictyum* 

The monophyly of *Cyrtomidictyum* was strongly supported by four *cp*DNA sequence data sets that include all four currently recognized species. In all phylogenetic analyses (Figs. 2, 4, 5), this genus was the basal-most lineage and clearly separated from other extant polystichoid taxa.

As a distinct genus, *Cyrtomidictyum* was proposed by Ching early in 1940 based on *Aspidium lepidocaulon* Hooker, a species originally known from Tsu-Shima Island, and later found to be rather widely distributed in South Korea, Japan and China (Ching 1940, 1957). *Cyrtomidic*-

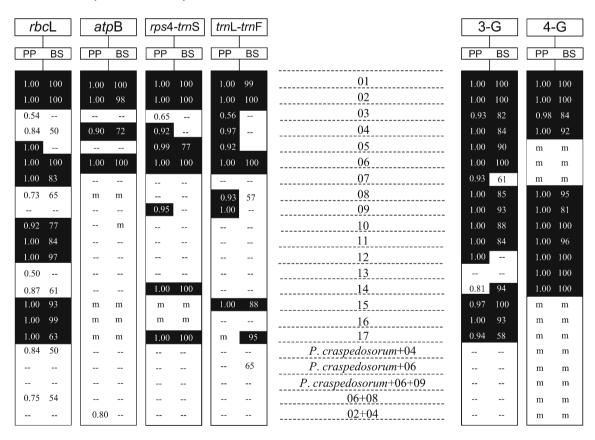
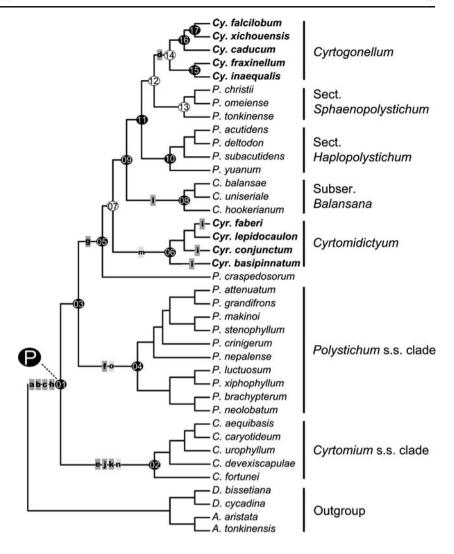


Fig. 3 Support values for major clades. Each row corresponds to a clade receiving either Bayesian posterior probability (PP)≥0.50 or Maximum Likelihood bootstrap support (BS)≥50% from single- or multiple-gene data sets. Clades 01–14 were resolved in the three-gene combined analysis and correspond to the numbered nodes in Figure 4. The remaining clades were supported in different single-gene data sets; their composition is indicated in terms of nodes resolved in the combined

analysis (Fig. 4) and/or by species name (where necessary). For each of the six data matrices (framed columns; 3-G=combined *rbc*L, *rps4-trn*S and *trnL-trn*F; 4-G=combined *rbc*L, *atp*B, *rps4-trn*S and *trnL-trn*F), PP and BS values≥50 are given. Black cells highlight values of PP≥ 0.90 and BS≥70. Four separate single-gene data sets differ in taxon sampling density due to missing sequence data (marked "m")



Fig. 4 50% majority rule consensus tree from Bayesian analyses of the three-gene (rbcL, rps4-trnS, and trnL-trnF) data set. Numbered solid circles indicate nodes receiving good support in both Bayesian (PP≥ 0.90) and Maximum Likelihood (BS≥70) analyses; open circles indicate nodes not receiving good support from either measure: see also Figure 3. Single letters in dark and light grey squares on branches refer to synapomorphic indels in the rps4-trnS and trnL-trnF alignments, respectively. "P" denotes polystichoid ferns; major groups also indicated at right



tyum has been regarded as a member of Polystichum, with which it shares features such as once-pinnate leaves, auriculate pinnae and free veins. Unique to Cyrtomidictyum are the small, generally biseriate and exindusiate sori which are dorsal on veinlets, the entire, non-aristate-serrate margin of the pinnae, uniform or sub-dimorphic leaves, and the leaf apices which often are elongated into a scaly whip-like and radicant stolon. In *Polystichum* the sori are dorsal, terminal or submarginal with a round and entire indusium which is peltately attached, the margins of pinnae are aristate to serrate, and the leaves are usually monomorphic. These diagnostic character states, which characterize Cyrtomidictyum as a natural group, also occur in some species of Polystichum. For example, members of sections Mastigopteris, Stenopolystichum and Macropolystichum of Polystichum sometimes have elongated rachis apices without pinnae or with one to several gammae on the rachis. Individual character states occur sporadically in some species, but are never found in this combination in any single species of Polystichum, nor are they as clearly

developed as in *Cyrtomidictyum*. Consequently, *Cyrtomidictyum* is morphologically well-defined and has been recognized as a natural group by several authors (Ching 1978; Kung et al. 2001; Pichi Sermolli 1977; Tryon and Tryon 1982; Wu and Ching 1991).

Using *rbc*L sequence data, Little and Barrington (2003) supported the separation of *Cyrtomidictyum* as a genus distinct from *Polystichum* s.l.. The authors also argued for the inclusion of *P. tripteron* and *P. deltodon* in *Cyrtomidictyum*. More broadly, Li et al. (2008) proposed to expand *Cyrtomidictyum* to include *Cyrtogonellum*, several *Polystichum* and *Cyrtomium* species, namely the CCPC clade. However, both these suggestions were not fully supported in our study. All four *Cyrtomidictyum* species fell into a well-supported clade which was clearly separated from other polystichoid ferns. The monophyly of *Cyrtomidictyum* and its separation from other polystichoid ferns was supported not only by the accumulation of substitutions, but also by the occurrence of one unique deletion of 15 base pairs (TTAGCTAGATTCCGA) in the *rps4-trnS* 



**Table 2** Synapomorphic indels revealed by the alignment of the two intergenic spacer sequences, *rps*4-*trn*S and *trn*L-*trn*F

Indels	Sequences
rps4-trnS alignment	
Insertions	
a	TTTA
b	TCCRGCCCGK
c	GRARAAATKAATC
d	TAATTCGTACCGAGGC
e	AT
f	TCTCT
g	TCTCT
Deletions	
h	CRA
i	TTAGCTAGATTCCGA
j	GTCTCTG
k	117 base pairs
1	TTGGATGACTTG
trnL-trnF alignment	
Insertions	
m	AGT
n	CCCTTCTA
0	CATCAACGATTTGA

alignment in three *Cyrtomidictyum* species, though not in *C. lepidocaulon*.

Very low to no sequence divergence was found among the four species of *Cyrtomidictyum* for all four sequenced chloroplast genome regions. Only in the *rps4-trnS* data two subgroups were identified within the genus, consisting of *C. lepidocaulon* and *C. faberi*, and *C. basipinnatum* and *C. conjunctum*, respectively. The diversification of this clade and the taxonomic status of its species requires additional studies using more variable markers and including nuclear markers.

# Phylogenetic relationships of Cyrtogonellum

Cyrtogonellum is an eastern Asian endemic genus with restricted geographical distribution in southwest China, specifically on limestone substrates in Yunnan, Guizhou and adjacent regions (Kung et al. 2001). Its taxonomic affinity and phylogenetic position have been controversial, with some morphologists treating the name Cyrtogonellum as a synonym of Phanerophlebia, Cyrtomium or Polystichum (Copeland 1947; Kramer et al. 1990; Tryon and Lugardon 1991). As regards morphological characters, Cyrtogonellum shows many similarities to Polystichum. The latter is generally defined by a short and erect rhizome, once-pinnate leaves which are always ovate-oblong or lanceolate, stipes and rachises which are sulcate adaxially, entire or crenate-serrate pinnae, round sori on the apices of

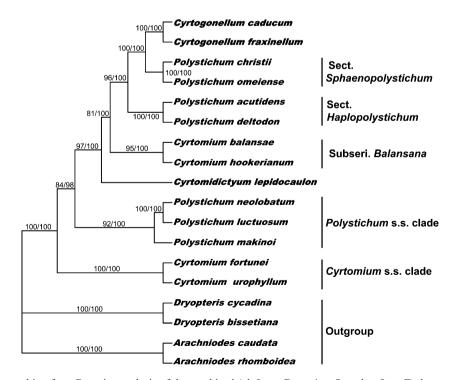


Fig. 5 Consensus tree resulting from Bayesian analysis of the combined (rbcL, atpB, rps4-trnS, and trnL-trnF) data set. Numbers on branches are Bayesian (PP)/Maximum Likelihood (BS) support values



veinlets, and entire, round to reniform indusia which are peltately attached. *Cyrtogonellum* shares all these features.

The molecular data, however, indicate different phylogenetic relationships. The patterns retrieved in our multiplegene analysis of a more extensive sample of Cyrtogonellum agreed with those found in previous analyses of Chinese Polystichum by Lu et al. (2007) and Li et al. (2008) in which Cyrtogonellum showed a close affinity to Cyrtomium subser. Balansana and members of three Polystichum sections, especially to sect. Sphaenopolystichum. The close relationship between Cyrtogonellum and Polystichum sect. Sphaenopolystichum found strong support in the DNA sequence variation, yet no obvious morphological and/or molecular synapomorphy has been detected. The most remarkable difference between Cyrtogonellum and sect. Sphaenopolystichum is that the latter has deeply divided lamina segments and often compound leaves whereas Cyrtogonellum has once-pinnate leaves. A conflict between morphological and molecular data was also found in Gymnogrammitis. This polygrammoid fern has long been associated to davallioids based on similarity in blade shape and dissection, yet molecular evidence demonstrated the inclusion of Gymnogrammitis in polygrammoid ferns (Schneider et al. 2002).

Although our sampling of *Cyrtogonellum* and its putative relatives did not provide final evidence for its systematic placement, our results allow an hypothesis on its taxonomic status. We suggest to transfer three sections of *Polystichum* (sect. *Haplopolystichum*, sect. *Sphaenopolystichum* and sect. *Crucifilix*) and *Cyrtomium* subser. *Balansana* to *Cyrtogonellum*, and to accept a new genus definition. This treatment would allow conservation of the genus *Cyrtogonellum* as currently defined. However, it is difficult to find morphological synapomorphies for this enigmatic group of polystichoids.

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### References

- Chen, Z. D., Wang, X. Q., Sun, H. Y., Han, Y., Zhang, Z. X., Zou, Y. P., et al. (1998). Systematic position of the Rhoipteleaceae: evidence from nucleotide sequences of the *rbcL* gene. *Acta Phytotaxonomica Sinica*, 36, 1–7.
- Ching, R. C. (1938). A revision of the Chinese and Sikkim-Himalayan Dryopteris with reference to some species from neighbouring

- regions. Bulletin of the Fan Memorial Institute of Biology/ Botanical Series, 8, 327–331.
- Ching, R. C. (1940). The studies of Chinese ferns—XXXIII. Bulletin of the Fan Memorial Institute of Biology/Botanical Series, 8, 173–184.
- Ching, R. C. (1957). Cyrtomidictyum Ching, a yet little known Chinese fern genus. Acta Phytotaxonomica Sinica, 6, 255–266.
- Ching, R. C. (1978). The Chinese fern families and genera: systematic arrangement and historical origin. *Acta Phytotaxonomica Sinica*, *16*(3), 1–19. 16(4), 16–37.
- Copeland, E. B. (1947). Genera filicum. The genera of ferns. Waltham: Chronia Botanica Company.
- Doyle, J. J., & Doyle, J. L. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin*, 19, 11–15.
- Gastony, G. J., & Rollo, D. R. (1995). Phylogeny and generic circumscriptions of cheilanthoid ferns (Pteridaceae: Cheilanthoideae) inferred from rbcL nucleotide sequences. American Fern Journal, 85, 341–360.
- Geiger, J. M., & Ranker, T. A. (2005). Molecular phylogenetics and historic biogeography of Hawaiian *Dryopteris* (Dryopteridaceae). *Molecular Phylogenetics and Evolution*, 34, 392–407.
- Guindon, S., & Gascuel, O. (2003). A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. Systematic Biology, 52, 696–704.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series, 41, 95–98.
- Hasebe, M., Wolf, P. G., Pryer, K. M., Ueda, K., Ito, M., Sano, R., et al. (1995). Fern phylogeny based on rbcL nucleotide sequences. American Fern Journal, 85, 134–181.
- Kramer, K. U., Holttum, R. E., Moran, R. C., & Smith, A. R. (1990).Dryopteridaceae. In K. Kubitzki (Ed.), *The families and genera of vascular plants* (pp. 99–144). Berlin: Springer-Verlag.
- Kung, H. S., Chu, W. M., He, Z. R., & Zhang, L. B. (2001).
  Polystichum. In H. S. Kung (Ed.), Flora reipublicae popularis sinicae (pp. 1–177). Beijing: Science.
- Li, C. X., & Lu, S. G. (2006). Phylogenetic analysis of Dryopteridaceae based on chloroplast rbcL sequences. Acta Phytotaxonomica Sinica, 44, 503–515.
- Li, C. X., Lu, S. G., & Yang, Q. (2004). Asian origin for *Polystichum* (Dryopteridaceae) based on *rbc*L sequences. *Chinese Science Bulletin*, 49, 1146–1150.
- Li, C. X., Lu, S. G., & Barrington, D. S. (2008). Phylogeny of Chinese *Polystichum* (Dryopteridaceae) based on chloroplast DNA sequence data (*trn*L-F and *rps*4-*trn*S). *Journal of Plant Research*, 121, 19–26.
- Little, D. P., & Barrington, D. S. (2003). Major evolutionary events in the origin and diversification of the fern genus *Polystichum* (Dryopteridaceae). *American Journal of Botany*, 90, 508–514.
- Liu, H. M., Zhang, X. C., Chen, Z. D., & Qiu, Y. L. (2007a). Inclusion of the eastern Asia endemic genus Sorolepidium in Polystichum (Dryopteridaceae): evidence from the chloroplast rbcL gene and morphological characteristics. Chinese Science Bulletin, 52, 631– 638.
- Liu, H. M., Zhang, X. C., Wang, W., Qiu, Y. L., & Chen, Z. D. (2007b). Molecular phylogeny of the fern family Dryopteridaceae inferred from chloroplast *rbc*L and *atpB* genes. *International Journal of Plant Sciences*, 689, 1311–1323.
- Lu, J. M., Li, D. Z., Gao, L. M., Cheng, X., & Wu, D. (2005). Paraphyly of *Cyrtomium* (Dryopteridaceae): evidence from *rbcL* and *trnL-F* sequences data. *Journal of Plant Research*, 118, 129–135.
- Lu, J. M., Barrington, D. S., & Li, D. Z. (2007). Molecular phylogeny of the polystichoid ferns in Asia based on rbcL sequences. Systematic Botany, 32, 26–33.



Pichi Sermolli, R. E. G. (1977). Tentamen pteridophytorum genera in taxonomicum ordinem redigendi. *Webbia*, 31, 313–512.

- Posada, D., & Crandall, K. A. (1998). Modeltest: testing the model of DNA substitution. *Bioinformatics*, 14, 817–818.
- Ronquist, F., & Huelsenbeck, J. P. (2003). MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics*, 19, 1572–1574.
- Schneider, H., Smith, A. R., Cranfill, R., Haufler, C. H., Ranker, T. A., & Hildebrand, T. (2002). Gymnogrammitis dareiformis is a polygrammoid fern (Polypodiaceae)—resolving an apparent conflict between morphological and molecular data. Plant Systematics and Evolution, 234, 121–136.
- Taberlet, P., Gielly, L., Pautou, G., & Bouvet, J. (1991). Universal primers for amplification of three non-coding regions of chloroplast DNA. *Plant Molecular Biology*, 17, 1105–1109.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., & Higgins, D. G. (1997). The Clustal X windows interface: flexible strategies for multiple sequence alignment aided by

- quality analysis tools. *Nucleic Acids Research*, 24, 4876–4882.
- Thomson, J. A., Chikuni, A. C., & McMaster, C. S. (2005). The taxonomic status and relationships of bracken ferns (*Pteridium*: Dennstaedtiaceae) from sub-Saharan Africa. *Botanical Journal of the Linnean Society*, 148, 311–321.
- Tryon, A. F., & Lugardon, B. (1991). Spores of the Pteridophyta. New York: Springer-Verlag.
- Tryon, R. M., & Tryon, A. F. (1982). Ferns and allied plants, with special reference to tropical America. New York: Springer-Verlag.
- Wu, S. H., & Ching, R. C. (1991). Fern families and genera of China. Beijing: Science.
- Wu, S. G., Xiang, J. Y., & Phan, K. L. (2005). Some new records of ferns from Vietnam (1). Acta Botanica Yunnanica, 27, 249– 252
- Xie, Y. T. (1990). Classification and distribution of Cyrtogonellum Ching. Bulletin of Botanical Research, 10, 93–97.

